

PRIMARY IMMUNODEFICIENCY (PID) REQUEST FORM



PRIMARY IMMUNODEFICIENCY UNIT
Allergy & Immunology Research Centre (AIRC)
Institute for Medical Research (IMR)
National Institutes of Health (NIH)
No 1, Jalan Setia Murni U13/52
Seksyen U13 Setia Alam, 40170 Shah Alam, Selangor
Telephone: 03-3362 8386

For IMR Lab use only:

1. PATIENT DETAILS

Appointment Date given: _____

Name:		
NRIC:		RN:
DOB:	Age:	Clinic/Ward:
Gender:	Ethnicity:	Hospital:
Requesting Specialist:		
Requesting Specialist's Contact Number (<i>for notification of urgent results</i>):		

2. CLINICAL HISTORY (please add extra sheet if necessary)

3. FAMILY PEDIGREE (3 generations or more)

➤ Please send as a separate attachment (COMPULSORY)

4. INVESTIGATIONS REQUIRED (please tick)

* Cases must be discussed with PID unit officers

Preliminary testing		
i.	Lymphocyte subset enumeration (TBNK)	
Chronic Granulomatous Disease (CGD)*		
i.	Dihydrorhodamine (DHR) test	
Assessment of T cells function*		
i.	Lymphocyte activation test (LAT)	
X-linked agammaglobulinemia (XLA)*		
i.	Bruton tyrosine kinase (BTK) protein by flow cytometry	
ii.	BTK genetic testing	
Hemophagocytic lymphohistiocytosis (HLH)*		
i.	Serum soluble CD25 detection by ELISA	
Others: please specify*		
i.	Immunoglobulin G (IgG) subclass	

5. BLOOD COLLECTION DETAILS

Time:

Date:

FOR PID LABORATORY USE ONLY:

Healthy control received: ☐ Yes ☐ No ☐ N/A

6. APPLICANT DETAILS:

SAMPLE COLLECTION INSTRUCTIONS

1. All laboratory tests are performed on an appointment basis every working Monday – Thursday.
2. All tests **EXCEPT THE IgG SUBCLASS** require an appointment. Please call our unit at 03 – 3362 8386 to request an appointment.
3. Samples must reach the PID laboratory before 1 p.m. on the appointment date given.
4. Sample requirements for each test can be found in the table below.
5. For additional information regarding each test, please refer to the IMR test list. (<https://imr.nih.gov.my/testlist>)

Test	Vacutainer	Transportation temp	Sample volume	TAT*
TBNK	EDTA	Room temperature / Ambient	2 mL of fresh blood	5 days
<i>TBNK: measurement of total T cells (CD3), T helper cells (CD4), T cytotoxic cells (CD8), B cells (CD19) and Natural Killer cells using flow cytometry</i>				
DHR test	Lithium heparin	Room temperature / Ambient (STRICTLY NO ICE)	2 mL of fresh blood each from patient and unrelated healthy control	10 days
<i>DHR Test: Assessment of neutrophils respiratory burst using flow cytometry</i>				
LAT	EDTA	Room temperature / Ambient (STRICTLY NO ICE)	5 mL of fresh blood each from patient and unrelated healthy control	10 days
<i>LAT: Assessment of interferon-gamma production in T cells after stimulation with anti-CD3 and PHA</i>				
BTK protein by flow cytometry	EDTA	Room temperature / Ambient	2 mL of fresh blood each from patient, mother and unrelated healthy control	10 days
BTK genetics	EDTA	In Ice	10 mL of fresh blood each from patient and mother	4 months
<i>BTK flow: Assessment of BTK protein expression on monocytes using flow cytometry</i> <i>BTK genetics: Sequencing of the BTK gene</i> <i>(Indication: Male with B cells <2% + hypogammaglobulinemia)</i>				
sCD25	Plain tube	Room temperature / Ambient	5 mL of fresh blood	21 days
<i>sCD25: Quantitation of sCD25, a marker for hemophagocytic lymphohistiocytosis</i>				
IgG subclass	Plain tube	Room temperature / Ambient	5 mL of fresh blood	21 days
<i>IgG subclass: Quantitation of IgG1, IgG2, IgG3 and IgG4 using turbidimetry</i>				

Healthy control sample: Blood sample from any healthy individuals, unrelated to the patients. The sampling time for the control should be immediately before or after blood sampling of the patient.

*TAT: Turnaround time is subject to working days