## A COMMON FACTOR FOR THE SAMPLE THAT AFFECTS THE EXAMINATION'S PERFORMANCE IN DISEASE SECTION

UNIT	TEST	SAMPLE	FACTOR	CONSEQUENCES
Tibi/Kusta	MTB Identification and Drug susceptibility testing (DST)	Sputum	<ul> <li>Unsatisfactory sample (saliva)</li> <li>Sample delivery - ≥ 48 hours after sample collection</li> <li>Storage temperature - ≥ 2-8°C</li> </ul>	<ul><li>Low bacterial Load</li><li>normal flora overgrowth</li><li>normal flora overgrowth</li></ul>
		Gastric lavage	<ul> <li>No sodium bicarbonate added</li> <li>Storage temperature - ≥ 2-8°C</li> <li>Sample delivery - ≥4 hours after sample collection</li> </ul>	<ul> <li>acidic sample can inhibit bacterial growth</li> <li>acidic sample can inhibit bacterial growth</li> <li>acidic sample can inhibit bacterial growth</li> </ul>
	Line Probe Assay (LPA)	Sputum	<ul> <li>AFB direct smear &lt;1+</li> <li>Sample delivery - &gt; 48 hours after sample collection</li> <li>Storage temperature - &gt; 2-8°C</li> </ul>	<ul> <li>Low bacterial Load</li> <li>normal flora overgrowth</li> <li>normal flora overgrowth</li> </ul>
	PCR TB/NTM	Sputum	<ul> <li>Sample delivery - <u>&gt;</u> 48 hours after sample collection</li> <li>Storage temperature - <u>&gt;</u> 2-8°C</li> <li>Bloody sample</li> </ul>	<ul> <li>normal flora overgrowth</li> <li>normal flora overgrowth</li> <li>Result invalid -RBC as an inhibitor</li> </ul>
	Interferon- gamma release assays (IGRA)	Whole blood	<ul> <li>Insufficient sample – volume</li> <li>&lt;0.8 ml or &gt;1.2 ml</li> <li>Sample hemolyzed</li> <li>Sample lipemic</li> <li>Improper sample shaking</li> <li>Improper sample incubation</li> <li>Improper sample centrifugation</li> </ul>	Result invalid
	GeneXpert	Sputum	<ul> <li>Unsatisfactory sample (saliva)</li> <li>Sample delivery - ≥ 48 hours after sample collection</li> <li>Storage temperature - ≥ 2-8°C</li> <li>Sample from patients who have received no antituberculosis therapy, or less than 3 days of therapy in the last 6 months.</li> <li>Bloody sample - RBC as an inhibitor</li> </ul>	<ul> <li>Low bacterial Load</li> <li>normal flora overgrowth</li> <li>normal flora overgrowth</li> <li>Result interference (false result)</li> <li>Result invalid -RBC as an inhibitor</li> </ul>
		Gastric lavage	<ul> <li>Storage temperature - ≥ 2-8°C</li> <li>Sample delivery - ≥ 4 hours after sample collection</li> <li>Sample from patients who have received no antituberculosis therapy, or less than 3 days of therapy in the last 6 months.</li> </ul>	<ul> <li>acidic sample can inhibit bacterial grow</li> <li>acidic sample can inhibit bacterial grow</li> <li>Result interference (false result)</li> </ul>

Note: Refer to Disease Section Test Handbook for sample collection and criteria

Rev. no: 00 Effective Date: 16/5/2025

UNIT	TEST	SAMPLE	FACTOR	CONSEQUENCES
Biokimia	TSH/FT4 Test Cholinesterase Test Liver Function Test Renal Profile Lipid Profile Miscellaneous Tests: -Calcium -Phosphorus	Serum /Plasma	<ul> <li>Inappropriate sample collection techniques for example drawing blood too quickly or leaving the tourniquet on for too long</li> <li>Improper order of draw</li> <li>Use of incorrect collection tubes</li> <li>Insufficient sample volume</li> <li>The collection site is not properly disinfected or if the collection tubes are contaminated.</li> <li>Inappropriate use of anticoagulants/ Improper mix</li> </ul>	Hemolysis (breakdown of red blood cells)      Increase possibility of the carryover of additive contamination     Inaccurate result/Hemolysis      Inaccurate result     Contamination of the sample      Clotting
	-Pnospnorus -Fasting Blood Glucose -Random Blood Glucose		of anticoagulant with sample or improper handling of blood  Delay in transportation of samples  Improper storage temperature of samples  Improper & delay of sample centrifugation/ separation  Not fasting prior to sample collection of Fasting Blood Glucose Test & Lipid Profile	<ul> <li>Degradation of sample         /Inaccurate result</li> <li>Degradation of sample         /Inaccurate result</li> <li>Degradation of sample         /Inaccurate result</li> <li>Inaccurate result</li> </ul>
	Adenosine Deaminase Test	Pleural fluid	<ul> <li>Incorrect sample collection technique (Aseptic technique during thoracentesis is required).</li> <li>Puncture Accidents-Accidental punctures during thoracentesis</li> <li>Insufficient sample volume</li> <li>Improper use of collection tube</li> <li>Delay in transportation of samples</li> <li>Improper storage temperature of samples</li> <li>Improper &amp; delay of sample centrifugation/ separation</li> </ul>	<ul> <li>Contamination of sample</li> <li>Can introduce blood into the sample</li> <li>Inaccurate result</li> <li>Inaccurate result</li> <li>Degradation of sample /Inaccurate result</li> </ul>
Bakteriologi	Culture test	Rectal swab/ Stool swab/ Throat swab/ Nasal swab/ Nasopharyngeal swab/ Pernasal swab	<ul> <li>Improper storage temperature of samples (no ice)</li> <li>Delay in shipment (Sample delivery &gt; 6 hours after sample collection</li> <li>Wrong transport media</li> <li>Wrong swab</li> </ul>	<ul> <li>Overgrowth of Normal flora</li> <li>Low yield of pathogen &amp;         Overgrowth of Normal flora</li> <li>No yield of pathogen</li> <li>No yield of pathogen</li> </ul>
	Leptospira MAT	Serum	<ul> <li>Improper &amp; delay of sample centrifugation/ separation</li> <li>Insufficient sample volume</li> </ul>	<ul> <li>Inaccurate result/ unable to see reaction with Leptospira serovar</li> <li>Unable to process sample</li> </ul>
	Leptospira PCR	Blood in EDTA	<ul><li>Sent serum sample</li><li>Insufficient sample volume</li></ul>	Leptospira DNA Not     Detected
	Syphilis Rechecking	Serum	<ul> <li>Insufficient sample volume</li> <li>Improper &amp; delay of sample centrifugation/ separation</li> <li>Lysed serum</li> <li>Insufficient sample volume</li> </ul>	Inaccurate result / unable to see agglutination

Note: Refer to Disease Section Test Handbook for sample collection and criteria

Rev. no: 00 Effective Date: 16/5/2025

UNIT	TEST	SAMPLE	FACTOR	CONSEQUENCES
Virology	Chikungunya/ Dengue/ Flavivirus/Zika Virus PCR	Serum /Blood	<ul> <li>Inadequate Sampel volume</li> <li>Sample not collected within specific days from onset of illness</li> <li>Storage temperature:&gt; 8°C</li> </ul>	<ul> <li>Inadequate sample size will result in template DNA present in the reaction leading to low yield of PCR product</li> <li>Decline in viral load</li> <li>Nucleic acids degradation</li> </ul>
	Enterovirus PCR –HFMD/ Conjunctivitis Respiratory	Mouth Ulcer Swab/ Vesicle Swab/ Rectal Swab/ Throat Swab/ Stool/	<ul> <li>Sample taken using other than flocked or dacron swab in VTM (volume 2-2.5ml) or sterile container.</li> <li>Sample not collected within specific days from onset of</li> </ul>	<ul> <li>Flocked and Dacron swabs are non-inhibitory to PCR</li> <li>Decline in viral load</li> </ul>
	Virus PCR  (Influenza A H1N1, H3N2, Influenza B, Adenovirus, RSV, SARS- CoV-2, MERS- CoV)  Monkeypox Virus PCR	Pleural fluid / Eye swab (for Conjunctivitis) Nasopharyngeal Swab/ Nasal swab/ Throat Swab/ combined OPS/NPS  Lesion fluid swab /Scab or crust Tonsillar swab/ Nasopharyngeal swab	illness  ◆ Storage temperature:> 8°C	Nucleic acids degradation
	Rotavirus	Fresh stool	<ul> <li>Sample not collected within specific days from onset of illness</li> <li>Sent in unsuitable storage temperature</li> </ul>	Decline in viral load
	Hepatitis B surface antigen or antibody / Rubella /Measles IgM	Serum/Blood	<ul> <li>Sampel volume: &lt;1ml</li> <li>Lysed serum/Plasma</li> <li>Unsuitable storage temperature:8°C (&lt;24hours) OR &gt;-70°C (&gt;48 hours)</li> </ul>	<ul> <li>Inadequate sample for testing</li> <li>False positive results</li> <li>Degradation of sample</li> <li>Inaccurate result</li> </ul>
Parasite	PCR Malaria	Blood in EDTA	Sample insufficient     Send plasma or serum instead     of EDTA	<ul> <li>Inadequate sample for testing</li> <li>Inappropriate sample will lead to poor yield of DNA and lead to false negative result.</li> </ul>
	Slide BFMP	Slides	Inappropriate packaging of slides  or cample collection and criteria.	Potential of broken slides and unable to proceed with rechecking.

Note: Refer to Disease Section Test Handbook for sample collection and criteria

Rev. no: 00 Effective Date: 16/5/2025